

## MHC Dextramer® Staining Protocol

**Intended use** Staining of antigen-specific T cells using one or more fluorochrome-labelled MHC Dextramer® reagents in one sample.

**Optimized for** Cat No. Wxxxxxx/Fxxxxxx/XDxxxxx  
MHC I Dextramer®, MHC II Dextramer®, and CD1d Dextramer®

**Materials Provided** MHC Dextramer® PE, APC and/or FITC

**Materials Required (not provided)** 4 mL Falcon disposable 12 x 75-mm test tubes or equivalent  
LoBind® Eppendorf tubes or equivalent  
Stain and wash buffer: PBS, 1-5% FCS, pH 7.4  
100 µM d-Biotin (e.g. Avidity, cat# BIO200) diluted in PBS, pH 7.4  
10x PBS, pH 7.4  
Antibodies identifying relevant surface markers (e.g., CD3, CD4, CD8 and optionally other desired antibodies and live-dead dye<sup>A</sup>). See the FAQ on [immudex.com](http://immudex.com) regarding [recommended antibody clones](#). The optimal choice of fluorochromes depends on the flow cytometer and experimental setup.

- Procedure**
1. Thaw and prepare PBMCs<sup>B</sup> and resuspend 1-3 x 10<sup>6</sup> PBMCs (for clonal cells, use 2-5 x 10<sup>4</sup> instead) in 50 µL stain and wash buffer.
  2. To prepare a pool of multiple MHC Dextramer® reagents (*calculation example can be found in Appendix 1*), mix the following reagents in an empty 1.5 mL LoBind® Eppendorf tube<sup>C</sup>:
    - a. Add 0.2 µL of 100 µM d-Biotin<sup>D</sup> per MHC Dextramer® reagent.
    - b. Add 10 µL of each MHC Dextramer® reagent.
    - c. Add 0.6 µL of 10x PBS<sup>D</sup> per MHC Dextramer® reagent.

*NB: When staining with a single MHC Dextramer® reagent, a and c can be omitted.*
  3. Vortex the MHC Dextramer® pool briefly. The MHC Dextramer® pool must be used directly after preparation and cannot be stored.
  4. Centrifuge the pool at 10.000 x g for 1 min. to avoid transferring any potential precipitate.
  5. Add the MHC Dextramer® pool to the cell sample and vortex briefly.
  6. Incubate in the dark at room temperature:
    - a. MHC I Dextramer® pool: 10 min. incubation<sup>E</sup>.
    - b. MHC II Dextramer® pool: 30 min. incubation<sup>E</sup>.
    - c. MHC I and II Dextramer® pool: 30 min. incubation<sup>E</sup>.
  7. Add relevant antibodies in the volume/concentration according to manufacturer's instructions:
    - a. If staining with MHC I Dextramer® reagents, use anti-CD8, anti-CD3, and optionally other phenotype markers.
    - b. If staining with MHC II Dextramer® reagents, use anti-CD4, anti-CD3, and optionally other phenotype markers.
  8. Incubate at room temperature in the dark for 20 min.

9. Wash cells by adding 2 mL stain and wash buffer. Centrifuge at 300 x g for 5 min. and remove the supernatant. Repeat washing for a total of 2 washes<sup>F</sup>.
10. Resuspend the pellet in desired volume of stain and wash buffer suitable for your flow cytometer.
11. Proceed to analyze the samples on a flow cytometer or store at 2-8 °C in the dark. For optimal results, do not store the samples longer than 2 hours before acquisition. Alternatively, fixed cells<sup>G</sup> can be stored at 2-8C in dark for up to 24 hours.

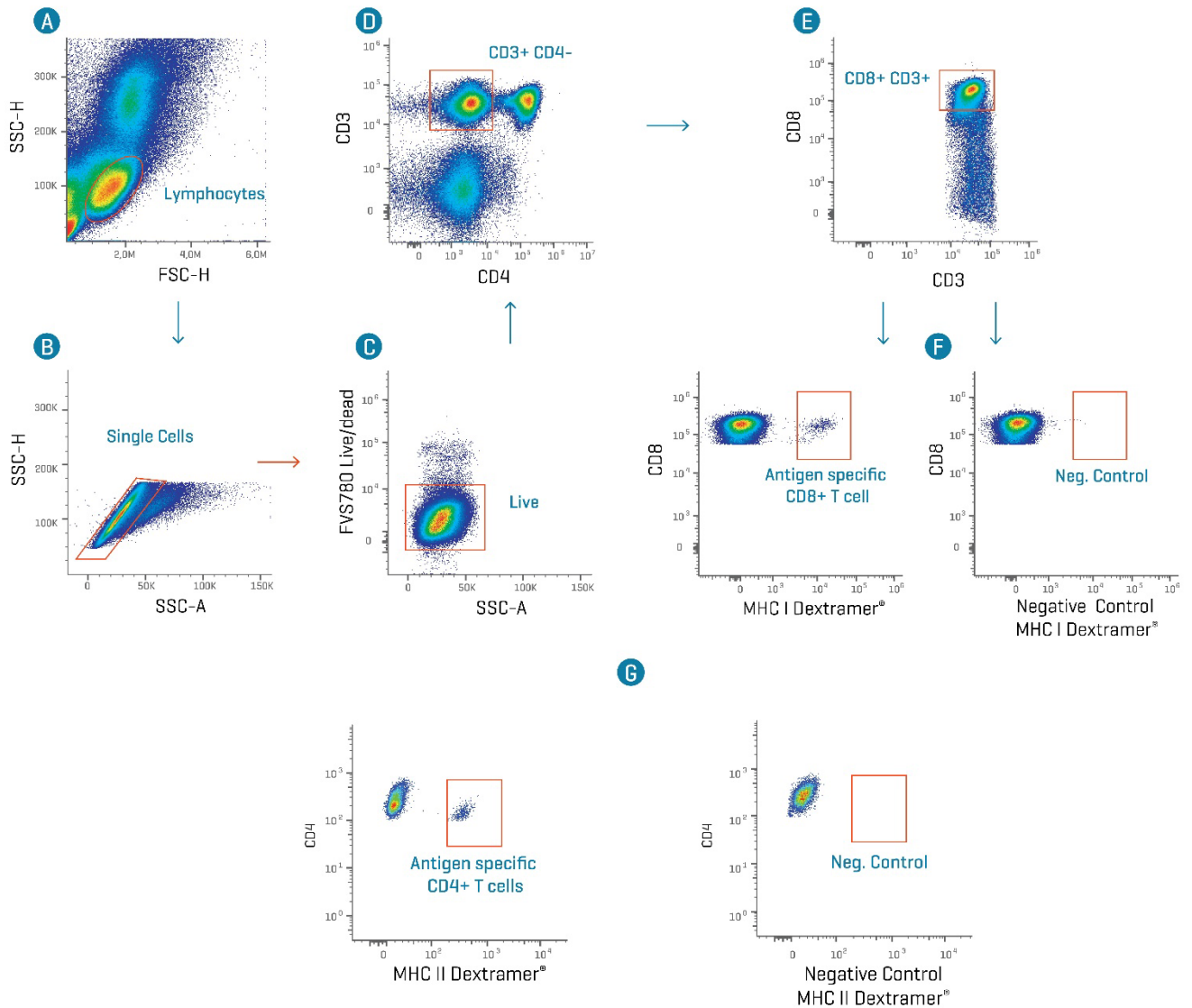
**Procedural notes**

- A. Live-dead staining can be performed at the beginning or end of staining procedure according to manufacturer's instructions.
- B. MHC Dextramer<sup>®</sup> staining can be performed on any cell suspensions, cell lines, TILs, or whole blood, if the cells are non-fixed. For whole-blood samples, stain with MHC Dextramer<sup>®</sup> before Red Blood Cell (RBC) lysis or use non-fixable RBC lysing solution.
- C. Always keep MHC Dextramer<sup>®</sup> reagents stored at 2-8 °C in the dark – the plastic vial only partially protects the reagents against light.
- D. d-biotin is required to avoid artefacts in the staining. 10x PBS will balance the salt concentration of the pool.
- E. Incubation time may be increased when using a high number of reagents in pool staining and requires optimization.
- F. Staining can be performed using 96-well microtiter plates. In that case after antibody incubation make 4 sequential washes using 200 µL stain and wash buffer per well. Centrifuge at 300 x g for 5 min. between each wash and remove supernatant.
- G. MHC Dextramer<sup>®</sup> stained cells can be fixed using 2% Methanol free formalin in PBS. Fixed samples may be washed and resuspended in stain and wash buffer prior to acquisition on a flow cytometer.

**Technical support**

For additional Tips & Tricks, FAQs and protocols, please visit <https://www.immudex.com/resources/> or contact our support team at [customer@immudex.com](mailto:customer@immudex.com)  
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**Analysis  
Guidelines**



**Fig. 1: Flow cytometry gating strategy using MHC I Dextramer<sup>®</sup> to identify antigen specific T-cells from samples of thawed hPBMCs. (A-F) gating of CD8+ antigen specific T cells. (A)** Lymphocytes were identified based on the forward (FSC) - and side scatter (SSC) profiles. **(B)** Next, doublets were excluded by gating the single cells in a side scatter height (SSC-H) & side scatter area (SSC-A) profile plot. **(C)** Dead cells were excluded according to the live-dead stain (FVS780), and the live cells were gated for further characterization. **(D)** To exclude CD4<sup>+</sup> T cells and Natural killer cells (NK) (positive for CD8 but not CD3), the CD3<sup>+</sup>/CD4<sup>-</sup> population was gated. **(E)** The CD3<sup>+</sup>/CD8<sup>+</sup> T cells were then gated, and **(F)** subsequently, the antigen-specific population of cells were determined by comparing the results of gating the MHC I Dextramer<sup>®</sup> labeled or MHC I Dextramer<sup>®</sup> Negative Control labeled cells. **(G)** Flow cytometry plots showing CD4<sup>+</sup> T helper cells labeled with MHC II Dextramer<sup>®</sup> or Negative Control MHC II Dextramer<sup>®</sup>.

## Appendix 1 Calculation Examples

Preparation of pools of MHC Dextramer<sup>®</sup> reagents for staining 1 sample:

Examples	100 µM d-Biotin	Total MHC Dextramer <sup>®</sup> Reagents	10x PBS	Total Volume
Per each MHC Dextramer <sup>®</sup>	0.2 µL	10 µL per MHC Dextramer <sup>®</sup>	0.6 µL	10.8 µL
2 MHC Dextramer <sup>®</sup> reagents	0.4 µL	20 µL MHC Dextramer <sup>®</sup>	1.2 µL	21.6 µL
3 MHC Dextramer <sup>®</sup> reagents	0.6 µL	30 µL MHC Dextramer <sup>®</sup>	1.8 µL	32.4 µL
10 MHC Dextramer <sup>®</sup> reagents	2 µL	100 µL MHC Dextramer <sup>®</sup>	6 µL	108 µL

Preparation of pools of MHC Dextramer<sup>®</sup> reagents for staining 2 samples:

*Note: When preparing a pool for more than 1 sample, we recommend preparing 20% overage of the pool, which is included in the examples below.*

Examples	100 µM d-Biotin	Total MHC Dextramer <sup>®</sup> Reagents	10x PBS	Total Volume
Per each MHC Dextramer <sup>®</sup>	0.2 µL	12 µL per MHC Dextramer <sup>®</sup>	0.7 µL	12.9 µL
2 MHC Dextramer <sup>®</sup> reagents	0.5 µL	24 µL MHC Dextramer <sup>®</sup>	1.4 µL	25.9 µL
3 MHC Dextramer <sup>®</sup> reagents	0.7 µL	36 µL MHC Dextramer <sup>®</sup>	2.2 µL	38.9 µL
10 MHC Dextramer <sup>®</sup> reagents	2.4 µL	120 µL MHC Dextramer <sup>®</sup>	7.2 µL	129.6 µL